

Protective roles of herbal teas against the toxicity of Bisphenol A on morphometry of uterus and improving murine reproductive health

Rasha Al-Eisa¹, Fatin Alsalmi¹, Rowa Kamal Zarah¹, Hamida Hamdi¹, Rokayya Sami^{2*}, Suzan Abushal³, Naseh Ateahallah Algehainy⁴, Mohammad Alanazi⁴, Faris Tayeb⁴, Ahlam Abbas Harasani⁵, Awatif Musallam Almehmadi⁶, Noor Allehyani⁷ and Abeer Ahmed Abu-zaid⁸

¹Department of Biology, College of Sciences, Taif University, Taif, Saudi Arabia

²Department of Food Science and Nutrition, College of Sciences, Taif University, Taif, Saudi Arabia

³Program of Food Sciences and Nutrition, Turabah University College, Taif University, Taif, Saudi Arabia

⁴Department of Medical Laboratory Technology, Faculty of Applied Medical Sciences, University of Tabuk, Tabuk, Saudi Arabia

⁵Department of Biological Sciences, College of Science, University of Jeddah, Jeddah, Saudi Arabia

⁶Department of Clinical Nutrition, Faculty of Applied Medical Sciences, Umm Al-Qura University, Makkah, Saudi Arabia

⁷Department of Medical Laboratories, College of Applied Medical Sciences, Shaqra University, Shaqra, Saudi Arabia

⁸Department of Food Science and Nutrition, Alkhurmah University College, Taif University, Taif, Saudi Arabia

Abstract: The purpose of the current study was to investigate the potential ameliorating murine reproductive effects of herbal tea extracts against bisphenol A-induced (BPA) cytotoxicity. A comparative study was applied among red, green, and blue teas in mice groups. Samples were coded as RTE, GTE and BTE groups, respectively. Several evaluations of murine reproductive were detected, such as diameters of uterine layers, pregnancy percentage, uterine, and offspring weights. The levels of malondialdehyde (MDA), oxytocin and cortisol were detected in plasma. The pregnancy percentage in the BTE + BPA group was 100% in the negative control group. The uterine weights of the mice groups with herbal teas ranged from 4.12g to 4.77 g. The offspring weights of litter from dams after exposure to BPA and various herbal teas ranged from 17.53g to 25.12 g and from 20.01 g to 28.19 g for females and males, respectively. The BPA toxicity exposure increased MDA levels to reach 19.18ng/ml in the BTE + BPA group as compared with the negative control group of 14.32ng/ml. The exposure to BPA toxins decreased the oxytocin level in the negative control group from 32.55ng/ml to 25.55ng/ml in the RTE + BPA group, while the cortisol levels reached 31.39ng/ml in the GTE + BPA group. The appearance of the luminal epithelial cells and endometrium in the BTE + BPA group was close to normal. It was indicated that herbal teas, particularly blue tea can be able to improve murine reproduction, while the harmful effects of BPA cannot entirely be mitigated.

Keywords: Toxicity, bisphenol A, uterus, mice, improving murine reproductive.

Submitted on 03-06-2024 – Revised on 09-07-2024 – Accepted on 25-07-2024

INTRODUCTION

Tea is considered as the 2nd beverage after water. Herbal teas have become more widespread for avoiding the harmful effects of caffeinated drinks (Lakshan *et al.*, 2019). Herbal teas can be made from several types of flowers such as the butterfly blue pea, which has more antioxidant activities than green tea to protect the body from the free radicals damage (Ramli *et al.*, 2021). Blue tea is a caffeine-free tea that can be consumed in dry or fresh leaves (Minh, 2020). Blue tea is rich in catechin components which help in burning belly fat and losing weight with a calming effect and stress-relieving properties (Escher *et al.*, 2020). The consumption of blue tea can boost metabolism and consume extra calories (Kaewmanee *et al.*, 2011). The actual teas belong to *Camellia sinensis* and can vary into different types such as green, black, red and white teas. Green tea is a non-

fermented tea that is well-known for its health benefits (Ohishi *et al.*, 2016). It can treat or even prevent several diseases, such as cancer, heart disease, diabetes and obesity (Chatterjee *et al.*, 2012). Green tea has plenty of essential components such as phenolics, flavonoids, and antioxidants which may positively affect the metabolic pathways (Maiti *et al.*, 2019). Fresh green tea contains multiple nutrients such as carbohydrates, protein, amino acids, minerals, vitamins, organic acids and small amounts of fat (2%) (Henning *et al.*, 2004). The catechin components in green tea are higher than in black tea which appeared on catechins, galliccatechins, epicatechins, epigallocatechins, epicatechin gallates and epigallocatechin gallates structures (Majumder *et al.*, 2022). The fully fermented process leads to the production of black or red teas with brothy and bitter tastes (Łuczaj and Skrzydlewska, 2005). Red tea is rich in alkaloids, flavonoids, carbohydrates, phenolic acids and amino acids (Vermeer *et al.*, 2008). It has multiple medical and pharmacological uses as it enhances brain

*Corresponding author: e-mail: rokayya.d@tu.edu.sa

functions, memory, learning, nervous system and controls the level of serotonin and dopamine hormones (Leung *et al.*, 2001). Bisphenol A (BPA) is a chemical component usually used in plastic production due to its good properties (hardness, clear color and lightness) and coating on the insides of cans (Eladak *et al.*, 2015). BPA can be easily detected in human organs such as the fetal liver, cord blood, breast milk, and placenta (Huo *et al.*, 2015). BPA influence oxidative stress and pathogenesis for human and animal health (Zhang *et al.*, 2011). BPA overexposure plays an essential role in several diseases, alters the functions of the endocrine system and affects reproductive processes (Cousins *et al.*, 2002). Antioxidants and reducing agents such as herbal teas can protect from BPA toxicity (Nam *et al.*, 2010).

Therefore, the current research work aimed to investigate the protective roles of herbal teas against the toxicity of bisphenol A on the morphometry of the uterus and improve murine reproduction in mice.

MATERIALS AND METHODS

Herbal tea samples

Herbal tea samples and corn oil were purchased from the local market in Taif, KSA. Different tea samples were blue tea (*Clitoria ternatea*) red and green teas (*Camellia sinensis assamica*). Sample extracts were coded as BTE, RTE and GTE, respectively.

Aqueous extraction

The extraction of powdered tea samples was carried out by soaking 15 g of each type for 5 minutes in 500mL of boiled deionized water. The extracts were centrifuged at 10000 rpm for 10 minutes after cooling, while the supernatants were collected, freeze-dried and stored at -18 °C before and after mice testing.

Ethics approval and consent to participate

In the current experimental work was carried out under the Ethics Committee Number (8808/9988) at 2024 by Gulf Countries Association of Sciences in Animal Experimentation.

Animal study

Fifty healthy mice (*Mus musculus*) were randomly purchased from the vivarium of the Animal House in Jeddah, KSA. Females were 4 weeks old and about 25g (n=30), while males were 8 weeks old and about 35g (n=20). Mice were reared and housed under controlled environmental circumstances in separate stainless-steel cages with wood bedding. The condition was 25°C and 70 % humidity with a light cycle of 12 hours dark and 12 hours light. The mice had free access to clean water and a basic diet of 500g per cage weekly. Mice were subjected to 1 week of acclimatization before the experimental research.

Research design

BPA with a purity of 97% was purchased from Saudi Chemical Company, Jeddah, KSA. Mice were classified into 5 sets (10 rats each) and administrated as the following pattern: Group 1= (100mg/kg RTE + 5mg/kg BPA), Group 2= (100mg/kg GTE + 5mg/kg BPA) and Group 3= (100mg/kg BTE + 5mg/kg BPA). While Group 5 presents the (Positive Group (+)), mice were fed with a basic diet + 5 mg/kg BPA. Group 6 presents the (Negative Group (-)), mice were fed with a basic diet + 0.2ml/kg corn oil as the vehicle (Tyl *et al.*, 2002). The dose of BPA was calculated as (mg/kg) per body weight daily towards the murine models for 6 weeks. Mice were daily administered via oral gavage as described by (Akingbemi *et al.*, 2004).

Evaluation of murine reproductive processes

In a 1:1 ratio, five treated female mice were chosen to mate with fertile male mice. For five days, the pairings were housed in the same cages, while the number of pregnant female mice was noted. The pregnancy percentage was calculated, while the offspring weight for males and females was recorded after the 19th-21st day of gestation phase (Goh *et al.*, 2021).

Sample collection and medical evaluations

Daily body weights for mice were noted during the experimental period. On the final day of the experiment, the mice were anesthetized under a 40 mg/kg anesthesia injection of sodium dipyrone followed by the addition of 200 mg/kg in the drinking water. Blood samples were withdrawn and neatly collected from the heart, while the plasma was detected by centrifuging at 2000 rpm for 10 minutes and stored at -20°C for further analysis (Draper *et al.*, 1993; Baazeem *et al.*, 2024). The levels of malondialdehyde (MDA), oxytocin and cortisol were detected by ELISA kits (Sigma, CA, USA) (Draper *et al.*, 1993; Zaid *et al.*, 2018). The mice were sacrificed by using ketamine, while the whole uteruses were harvested and immediately weighed for the uterotrophic response evaluations as luminal epithelial cells' height, the thicknesses of endometrium, myometrium and perimetrium. Histomorphometry was performed on tissue-processed uterine sections using an image analyzer (NIS-Elements, Nikon, Japan).

STATISTICAL ANALYSIS

One-way analysis of variance was used to analyze each of the chosen parameters and the Bonferroni test (multiple comparisons) was then applied with SPSS Statistics 20 (IBM Corporation, Armonk, New York, USA). The data was presented as mean ± SEM. P less than 0.05 was deemed significant.

RESULTS

Effect of herbal teas on pregnancy percentage

The study of harmful biological effects on the reproductive system is known as reproductive toxicology

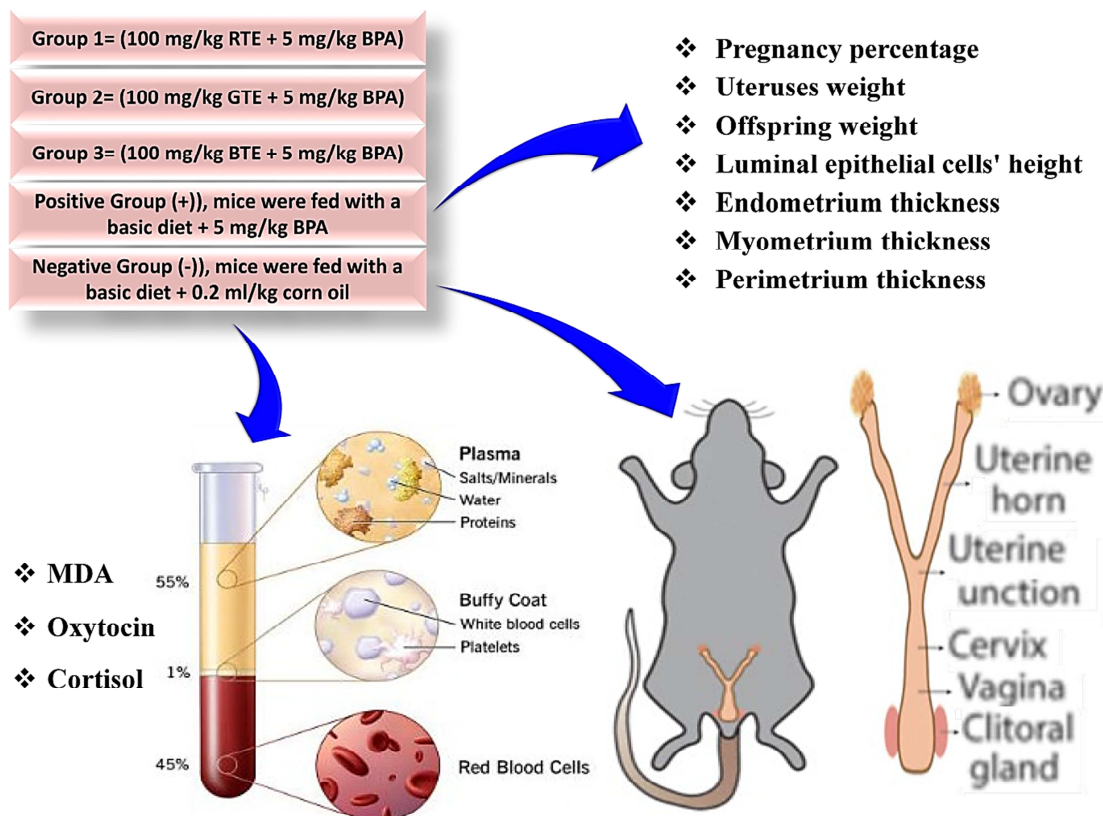


Fig. 1: Summary of the experimental processes

as a result of exposure to xeno-estrogenic substances such as BPA, which is present in infant formula and leaches from polycarbonate bottles (Biles *et al.*, 1997). As presented in fig. 2, changes in the pregnancy percentage were evaluated. The pregnancy percentage in the BTE + BPA group was 100% as the negative control group. BTE preserved the murine reproductive after exposure to BPA toxins compared with the RTE + BPA group and GTE + BPA group which reported 84.24 % and 97.59 %, respectively. Similar results were observed for the reduction in pregnancy percentage after exposure to BPA toxins (Ng *et al.*, 2021).

Effect of herbal teas on uterine weight

The effects of xeno-estrogen on the uterus are highly sensitive, particularly when an organ weight of $\geq 10\%$ is taken into consideration (Vigezzi *et al.*, 2015). Different trends were detected for uterine weights. When the mice were given BPA toxins concurrently, the increase in uterine weight gain was reduced. fig. 3. The positive control group's reported uterine weight was 5.83 g, while the negative control group's was 4.78 g. The uterine weights of the mice groups with herbal teas ranged from 4.12 g to 4.77 g. BTE + BPA group treatment prevented the effects of BPA toxins.

Effect of herbal teas on offspring weights

BPA exposure can have a detrimental effect on male mouse offspring's ability to produce sperm during a crucial developmental stage (Chitra *et al.*, 2003). Fig. 4 presented the offspring weights for both females and males. The offspring weights of litter from dams after exposure to BPA and various herbal teas ranged from 17.53g to 25.12g and from 20.01g to 28.19g for females and males, respectively. The negative control group reported 16.23g and 15.98g for females and males, respectively. Results were in agreement with Rubin *et al.*, (2001) who studied the effect of BPA on several parameters such as offspring weights and estrous cyclicity patterns.

Effects of herbal teas on plasma levels of MDA, oxytocin and cortisol

The effect of various herbal teas on plasma MDA levels is presented in fig. 5. The BPA toxicity exposure increased MDA levels to reach 19.18 ng/ml in the BTE + BPA group as compared with the negative control group 14.32 ng/ml. The RTE + BPA group reported the lowest level of

MDA 14.83 ng/ml. The findings were in link with Sangai *et al.* (2014) who investigated the BPA toxicity in a mice model's liver and kidney. Plasma oxytocin levels are presented in fig. 5. The exposure to BPA toxins decreased the oxytocin level in the negative control group from

32.55 ng/ml to 25.55 ng/ml in the RTE + BPA group. While the BTE + BPA group increased plasma oxytocin level to reach 28.73 ng/ml. The results agreed with Wu *et al.* (2012) who reported the functional uses of antioxidants to reduce oxytocin levels. The exposure to BPA toxins increased the cortisol levels in plasma especially in the positive control group to reach 39.01ng/ml, fig. 5. The phosphorylation of CAMP response element-binding protein may be enhanced by endometrium and perimetrium with an increase in the myometrium thickness (8.77µm, 180.91µm, 10.13µm and 255.10µm), respectively. The negative control group detected normal uterine diameters and healthy endometrial glands. The RTE + BPA group reported an increase in the myometrium thickness with a reduction in oestrogenic action, leading to a rise in cortisol levels (Lejonklou *et al.*, 2010). The administration of herbal teas decreased the cortisol levels to reach 31.39 ng/ml in the GTE + BPA group.

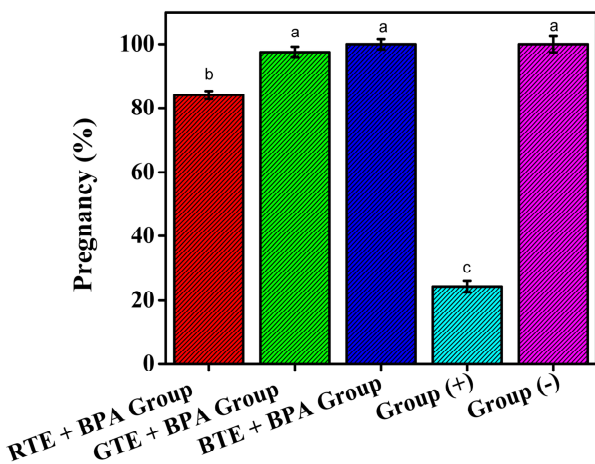


Fig. 2: Effect of Herbal Teas on Pregnancy Percentage among Mice Groups

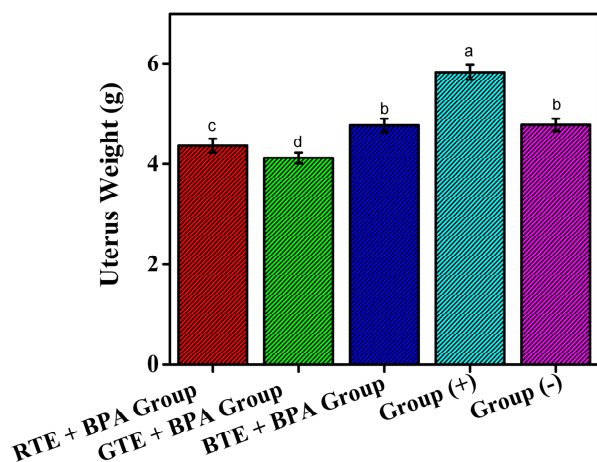


Fig. 3: Effect of Herbal Teas on Uterine Weight among Mice Groups

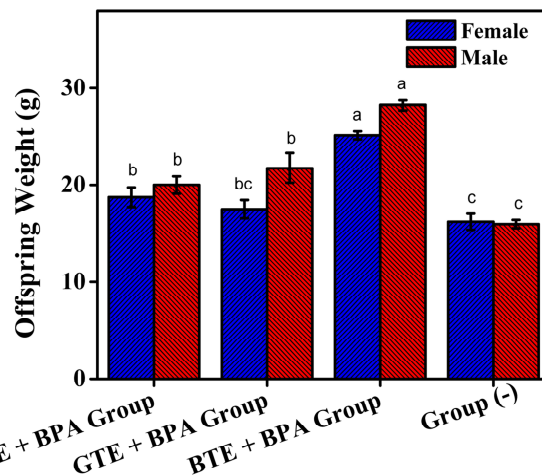


Fig. 4: Effect of Herbal Teas on Offspring Weights among Males and Females

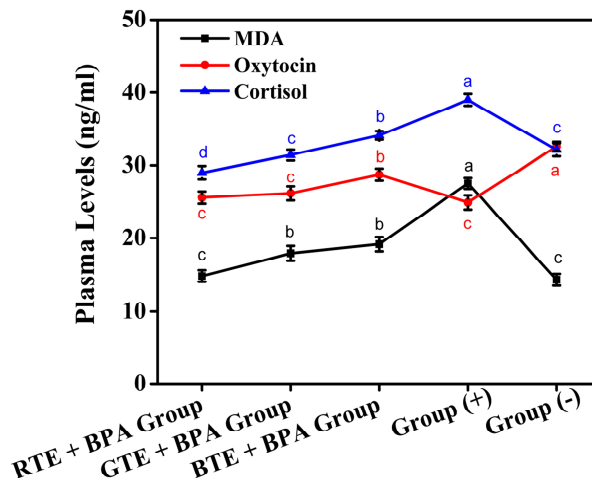


Fig. 5: Effect of Herbal Teas on Plasma levels (MDA, Oxytocin and Cortisol) among Mice Groups

Effect of herbal teas on the diameters of uterine layers

The effects of herbal teas on the diameters of uterine layers are presented in table 1. The positive control group exposed to BPA appeared unhealthy and shrunken resulting in a decrease in luminal epithelial cells' height and the thickness of various muscular uterine layers as luminal epithelial cells height and the thickness of endometrium and perimetrium (247.13µm 17.02µm 230.11µm and 12.59.10µm), respectively. Moreover, the appearance of the myometrium layer of RTE + BPA group was close to normal. The GTE + BPA group reported a reduction in myometrium thickness (233.95µm). BTE + BPA group reported an increase in luminal epithelial cells height and the thickness of endometrium and perimetrium (18.57µm, 244.11µm and 14.55µm), respectively. Moreover, the appearance of the luminal epithelial cells and endometrium for the BTE +

Table 1: Effect of herbal teas on the diameters of uterine layers among mice groups

	Endometrium thickness (μm)	Myometrium thickness (μm)	Perimetrium thickness (μm)	Luminal epithelial cells height (μm)
RTE + BPA Group	230.11 \pm 4.93 ^c	247.13 \pm 4.58 ^b	12.59 \pm 0.25 ^c	17.02 \pm 0.14 ^c
GTE + BPA Group	223.53 \pm 0.48 ^c	233.95 \pm 3.61 ^c	12.99 \pm 0.54 ^c	18.15 \pm 1.22 ^a
BTE + BPA Group	244.11 \pm 3.33 ^b	245.97 \pm 2.63 ^b	14.55 \pm 0.17 ^b	18.57 \pm 0.28 ^b
Group (+)	180.91 \pm 2.94 ^d	255.1 \pm 3.12 ^a	10.13 \pm 0.73 ^d	8.77 \pm 0.34 ^d
Group (-)	261.11 \pm 1.44 ^a	256.17 \pm 2.61 ^a	19.58 \pm 0.18 ^a	19.33 \pm 0.57 ^b

*Each value presents as mean \pm standard deviation. (n=3), Mean with different superscripts on the same column are significantly different ($p < 0.05$).

BPA group was close to normal. This indicated that the blue and red tea extracts might prevent negative changes in BPA-exposed mice. The results were in agreement with the previous findings by Markey *et al.* (2005).

DISCUSSION

It is commonly known that BPA is an endocrine disruptor which can mimic the effects of natural oestrogen, cause metabolic imbalances and develop illnesses (Newbold *et al.*, 2007). Prolonged exposure can result in irreversible infertility. In several animal models, exposure to BPA reduces fertility, messes with the estrous cycle, reduces the primordial follicle pool, and interferes with steroidogenesis (Ziv-Gal *et al.*, 2013). The findings on uterine weight were consistent with earlier research (Rodríguez *et al.*, 2010). Several factors can influence offspring weights as mice strains, age and the time of BPA exposure (Mendoza-Rodríguez *et al.*, 2011). The dietary intake of carotenoids in the diet can increase the production of male offspring than the females (McGraw *et al.*, 2005). This indicated that the blue tea extract might be employed as a treatment after BPA toxicity exposure and is safe for oral intake due to the presence of antioxidants. BPA is a xeno-estrogen that may lead to lipid peroxidation, all of which may cause organ diseases (Markey *et al.*, 2005). Moreover, epidemiological research revealed that infertile women had elevated levels of ROS (Polaka *et al.*, 2001). MDA can function as a co-carcinogen, inhibit protective enzymes, and cause hepatotoxicity (Bauer, 2000). According to a prior study (Patisaul *et al.*, 2012), the reduction levels of oxytocin in plasma were the reason why the adolescent animal model exposed to BPA toxics showed increased levels of anxiety. The hormone oxytocin aids in relaxation and lowers cortisol and blood pressure (Bhutada *et al.*, 2010). According to a study by Cheng and Li (2012), antioxidant components have been shown to inhibit the release of cortisol and corticotropin-releasing hormones under continuous stress. Bosquiazzo *et al.* (2013) reported the appearance of some deformities as persistent anoestrus which appeared in old animals undergoing reproductive senescence after exposure to BPA toxics.

CONCLUSION

BPA exposure negatively affects murine reproduction, but herbal teas have a protective effect, enhancing it. This

study investigated the potential benefits of three types of herbal teas: blue tea (*Clitoria ternatea*), red and green tea (*Camellia sinensis assamica*). All treatment groups showed improvements in uterine structure and weight; increased offspring weights and improved reproductive biomarkers, including plasma levels of MDA, oxytocin, and cortisol. The impact on other reproductive hormones and the specific mechanisms of BPA damage to other organs require further research.

ACKNOWLEDGMENT

The authors extend their appreciation to Taif University, Saudi Arabia for financial support to this work through Project No.TU-DSPP-2024-10.

REFERENCES

- Akingbemi BT, Sottas CM, Koulova AI, Klinefelter GR and Hardy MP (2004). Inhibition of testicular steroidogenesis by the xenoestrogen bisphenol A is associated with reduced pituitary luteinizing hormone secretion and decreased steroidogenic enzyme gene expression in rat Leydig cells. *Endocrinol.*, **145**(2): 592-603.
- Baazeem A, Helal M, Sami R, Alshehry G, Algarni E, Hilary U, Baakdah F, Alharthy SA and Mahmoud Johari D (2024). Positive effects of dietary honey and aflatoxin B1 on serum enzymes, superoxide dismutase activity, β -glucuronidase enzyme activity, and colonic probiotic bacteria on rats. *Mater. Express*, **14**(1): 60-65.
- Bauer G (2000). Reactive oxygen and nitrogen species: efficient, selective and interactive signals during intercellular induction of apoptosis. *Anticancer Res.*, **20**(6B): 4115-4139.
- Bhutada P, Mundhada Y, Bansod K, Ubgade A, Quazi M, Umathe S and Mundhada D (2010). Reversal by quercetin of corticotrophin releasing factor induced anxiety-and depression-like effect in mice. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry*, **34**(6): 955-960.
- Biles JE, McNeal TP and Begley TH (1997). Determination of bisphenol A migrating from epoxy can coatings to infant formula liquid concentrates. *J. Agric. Food Chem.*, **45**(12): 4697-4700.

- Bosquiazzo VL, Vigezzi L, Muñoz-de-Toro M and Luque EH (2013). Perinatal exposure to diethylstilbestrol alters the functional differentiation of the adult rat uterus. *J. Agric. Food Chem.*, **138**: 1-9.
- Chatterjee P, Chandra S, Dey P and Bhattacharya S (2012). Evaluation of anti-inflammatory effects of green tea and black tea: A comparative: *in vitro*: Study. *J. Adv. Pharm. Technol Res.*, **3**(2): 136-138.
- Cheng LC and Li LA (2012). Flavonoids exhibit diverse effects on CYP11B1 expression and cortisol synthesis. *Toxicol. Appl. Pharmacol.*, **258**(3): 343-350.
- Chitra KC, Rao KR and Mathur PP (2003). Effect of bisphenol A and co-administration of bisphenol A and vitamin C on epididymis of adult rats: A histological and biochemical study. *Asian J. Androl.*, **5**(3): 203-208.
- Cousins IT, Staples CA, Klečka GM and Mackay D (2002). A multimedia assessment of the environmental fate of bisphenol A. *Hum. Ecol. Risk Assess.*, **8**(5): 1107-1135.
- Draper HH, Squires EJ, Mahmoodi H, Wu J, Agarwal S, Hadley M (1993). A comparative evaluation of thiobarbituric acid methods for the determination of malondialdehyde in biological materials. *Free Radic. Biol. Med.*, **15**(4): 353-363.
- Eladak S, Grisin T, Moison D, Guerquin MJ, N'Tumba-Byn T, Pozzi-Gaudin S, Benachi A, Livera G, Rouiller-Fabre V and Habert R (2015). A new chapter in the bisphenol A story: Bisphenol S and bisphenol F are not safe alternatives to this compound. *Fertil. Steril.*, **103**(1): 11-21.
- Escher GB, Wen M, Zhang L, Rosso ND and Granato D (2020). Phenolic composition by UHPLC-Q-TOF-MS/MS and stability of anthocyanins from *Clitoria ternatea* L. (butterfly pea) blue petals. *Food Chem.*, **331**: 127341.
- Goh SE, Kwong PJ, Ng CL, Ng WJ and Ee KY (2021). Antioxidant-rich *Clitoria ternatea* L. flower and its benefits in improving murine reproductive performance. *Food Sci. Tech.*, **42**: e25921.
- Henning SM, Niu Y, Lee NH, Thames GD, Minutti RR, Wang H, Go VLW and Heber D (2004). Bioavailability and antioxidant activity of tea flavanols after consumption of green tea, black tea, or a green tea extract supplement. *Am. J. Clin. Nutr.*, **80**(6): 1558-1564.
- Huo X, Chen D, He Y, Zhu W, Zhou W and Zhang J (2015). Bisphenol-A and female infertility: A possible role of gene-environment interactions. *Int. J. Environ. Res. Public Health*, **12**(9): 11101-11116.
- Kaewmanee K, Priprem A and Preeprame S (2011). Stability and staining property of gel from roselle calyx extract and butterfly pea flower. *Planta Med.*, **77**(12): PK13.
- Lakshan SAT, Jayanath NY, Abeysekera WPKM and Abeysekera WKSM (2019). A commercial potential blue pea (*Clitoria ternatea* L.) flower extract incorporated beverage having functional properties. *Evid. Based Complementary Altern. Med.*, **20**: 2916914.
- Lejonklou MH, Hellman P, Botling J and Lind PM (2014). Bjorklund P Bisphenol A increases cortisol production by enhancing phosphorylation of CREB in normal human adrenocortical cells. S243-S257.
- Leung LaiKwok LL, Su YaLun SY, Chen RuoYun CR, Zhang ZeSheng ZZ, Huang Yu HY and Chen ZhenYu CZ (2001). Theaflavins in black tea and catechins in green tea are equally effective antioxidants. *J. Nutr.*, **131**(9): 2248-2251.
- Łuczaj W and Skrzydlewska E (2005). Antioxidative properties of black tea. *Prev. Med.*, **40**(6): 910-918.
- Maiti S, Nazmeen A, Medda N, Patra R and Ghosh TK (2019). Flavonoids green tea against oxidant stress and inflammation with related human diseases. *Clin. Nutr. Exp.*, **24**: 1-14.
- Majumder S, Das PC, Sami R, Ismail KA, Al-Mushhin AAM, Iqbal A, Ranganathan TV and Mazumder MAR (2022). Effect of spent green tea leaf extracts, butylated hydroxytoluene and repeated freezing-thawing on physicochemical and oxidative properties of chevon. *J. Biobased Mater. Bio.*, **16**(1): 56-67.
- Markey CM, Wadia PR, Rubin BS, Sonnenschein C and Soto AM (2005). Long-term effects of fetal exposure to low doses of the xenoestrogen bisphenol-A in the female mouse genital tract. *Biol. Reprod.*, **72**(6): 1344-1351.
- McGraw KJ, Adkins-Regan E and Parker RS (2005). Maternally derived carotenoid pigments affect offspring survival, sex ratio and sexual attractiveness in a colorful songbird. *Sci. Nat.*, **92**: 375-380.
- Melek ZOR, Sengul M, Karakutuk IA and Aksoy S (2023). Investigation about various infusion conditions on physical, chemical and antioxidant properties of *Clitoria ternatea* L. Tea. *J. Ins. Sci. Tech.*, **13**(3): 1738-1752.
- Mendoza-Rodríguez CA, García-Guzmán M, Baranda-Avila N, Morimoto S, Perrot-Applanat M and Cerbón M (2011). Administration of bisphenol A to dams during perinatal period modifies molecular and morphological reproductive parameters of the offspring. *Reprod. Toxicol.*, **31**(2): 177-183.
- Minh NP (2020). Efficacy of steaming, vacuum drying and stir-frying to total phenolic, flavonoid and organoleptic properties in butterfly pea flower (*Clitoria ternatea*) tea. *J. Entomol. Res.*, **44**(4): 621-624.
- Nam SH, Seo YM and Kim MG (2010). Bisphenol A migration from polycarbonate baby bottle with repeated use. *Chemosphere*, **79**(9): 949-952.
- Newbold RR, Jefferson WN and Padilla-Banks E (2007). Long-term adverse effects of neonatal exposure to bisphenol A on the murine female reproductive tract. *Reprod. Toxicol.*, **24**(2): 253-258.
- Ng CL, Tan GC, Yow YY, Gupta MK and Kwong PJ (2021). *Gracilaria changii* (Rhodophyta) alleviates bisphenol A-induced adverse reproductive

- abnormalities in mice. *Asian Pac. J. Trop. Med.*, **14**(1): 34-43.
- Ohishi T, Goto S, Monira P, Isemura M and Nakamura Y (2016). Anti-inflammatory action of green tea. *Antiinflamm. Antiallergy Agents Med. Chem.*, **15**(2): 74-90.
- Patisaul HB, Sullivan AW, Radford ME, Walker DM, Adewale HB, Winnik B, Coughlin JL, Buckley B and Gore AC (2012). Anxiogenic effects of developmental bisphenol A exposure are associated with gene expression changes in the juvenile rat amygdala and mitigated by soy. *Plos One*, **7**(9): e43890
- Polak G, Koziół-Montewka M, Gogacz M, Błaszowska I and Kotarski J (2001). Total antioxidant status of peritoneal fluid in infertile women. *Eur. J. Obstet. Gynecol. Reprod. Biol.*, **94**(2): 261-263.
- Ramli ME, Salleh RM, Tajarudin HA and Zulkurnain M (2021). Influence of amylose content on phenolics fortification of different rice varieties with butterfly pea (*Clitoria ternatea*) flower extract through parboiling. *LWT*, **147**: 111493.
- Rodríguez HA, Santambrosio N, Santamaría CG, Muñoz-de-Toro M and Luque EH (2010). Neonatal exposure to bisphenol A reduces the pool of primordial follicles in the rat ovary. *Reprod. Toxicol.*, **30**(4): 550-557.
- Rubin BS, Murray MK, Damassa DA, King JC and Soto AM (2001). Perinatal exposure to low doses of bisphenol A affects body weight, patterns of estrous cyclicity, and plasma LH levels. *Environ. Health Perspect.* **109**(7): 675-680.
- Sangai NP, Verma RJ and Trivedi MH (2014). Testing the efficacy of quercetin in mitigating bisphenol A toxicity in liver and kidney of mice. *Toxicol. Ind. Health.*, **30**(7): 581-597.
- Tyl RW, Myers CB, Marr MC, Thomas BF, Keimowitz AR, Brine DR, Veselica MM, Fail PA, Chang TY and Seely JC (2002). Three-generation reproductive toxicity study of dietary bisphenol A in CD Sprague-Dawley rats. *Toxicol. Sci.*, **68**(1): 121-146.
- Verma RJ and Sangai NP (2009). The ameliorative effect of black tea extract and quercetin on bisphenol A-induced cytotoxicity. *Acta Pol Pharm*, **66**(1): 41-44.
- Vermeer MA, Mulder TPJ and Molhuizen HOF (2008). Theaflavins from black tea, especially theaflavin-3-gallate, reduce the incorporation of cholesterol into mixed micelles. *J. Agric. Food Chem.*, **56**(24): 12031-12036.
- Vigezzi L, Bosquiazzo VL, Kass L, Ramos JG, Muñoz-de-Toro M and Luque EH (2015). Developmental exposure to bisphenol A alters the differentiation and functional response of the adult rat uterus to estrogen treatment. *Reprod. Toxicol.*, **52**: 83-92.
- Wu CH, Shieh TM, Wang KL, Huang TC and Hsia SM (2015). Quercetin, a main flavonoid in onion, inhibits the PGF2 α -induced uterine contraction *in vitro* and *in vivo*. *J. Funct. Foods*, **19**: 495-504.
- Zaid SSM, Othman S and Kassim NM (2018). Protective role of *Ficus deltoidea* against BPA-induced impairments of the follicular development, estrous cycle, gonadotropin and sex steroid hormones level of prepubertal rats. *J. Ovarian Res.*, **11**: 1-9.
- Zhang Z, Alomirah H, Cho HS, Li YF, Liao C, Minh TB, Mohd MA, Nakata H, Ren N and Kannan K (2011). Urinary bisphenol A concentrations and their implications for human exposure in several Asian countries. *Environ. Sci. Technol.*, **45**(16): 7044-7050.
- Ziv-Gal A, Craig ZR, Wang W and Flaws JA (2013). Bisphenol A inhibits cultured mouse ovarian follicle growth partially via the aryl hydrocarbon receptor signaling pathway. *Reprod. Toxicol.*, **42**: 58-67.